# **Protocol**

## **Detection of AP using the Alkaline Phosphatase Staining Kit II**

### **OVERVIEW**

The following procedure describes staining one well of a 6-well plate for Alkaline Phosphatase (AP) detection. For other plate formats, see the suggested amounts in Table 1.

| Product Description  | Cat. No. | Format  |
|--|----------|---------|
| Stemgent <sup>®</sup> Alkaline Phosphatase Staining Kit II | 00-0055  | 500 rxn |
| Components   | Size     | Storage |
| Fix Solution   | 25 ml    | 4°C     |
| AP Staining Solution A                                     | 10 ml    | 4°C     |
| AP Staining Solution B                                     | 10 ml    | 4°C     |
| AP Staining Solution C                                     | 10 ml    | 4°C     |

### ADDITIONAL MATERIALS REQUIRED

- 1X PBS
- Tween® 20
- Mounting medium (optional)
- 15 ml conical tubes

#### **MATERIAL PREPARATION**

#### PBST

In a 15 ml conical tube, add 10 ml of 1X PBS. Add 5  $\mu$ l of Tween 20 for a final concentration of 0.05%. Mix well and store at room temperature.

• AP Substrate Solution

For one well of a 6-well plate, mix 0.5 ml of Solution A and 0.5 ml of Solution B in a 15 ml conical tube. Incubate at room temperature for 2 minutes. Add 0.5 ml of Solution C.

**Note:** Prepare only the amount of AP Substrate Solution necessary for each experiment. Quantities can be scaled up or down, as long as a 1:1:1 ratio is preserved. For optimal results, the AP Substrate Solution should be **used within 30 minutes** after preparation. Discard any remaining solution.



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## **AP STAINING OF CELLS**

- 1. Aspirate the culture medium and wash the cells with 2 ml of 1X PBST.
- Add 1 ml of Fix Solution and incubate at room temperature for 2 to 5 minutes.
  Note: Do not over fix the cells. Excessive fixation will result in the loss of AP activity.
- **3.** Aspirate the Fix Solution and wash the fixed cells with 2 ml of 1X PBST. Do not allow the wells to dry.
- 4. Aspirate the 1X PBST and add 1.5 ml of freshly prepared AP Substrate Solution.
- 5. Incubate the cells in the dark (wrapped with foil or in a dark container) at room temperature for 5 to 15 minutes.

**Note:** Closely monitor the color change and stop the reaction when the color turns bright to avoid non-specific staining.

- **6.** Stop the reaction by aspirating the AP Substrate Solution and washing the wells twice with 2 ml of 1X PBS.
- 7. Cover the cells with 1X PBS or mounting medium to prevent drying.
- **8.** AP expression will result in a red or purple stain, while the absence of AP expression will result in no stain.
- 9. Store the plate at 4°C.

| Table 1. Suggested Amounts per Well |                      |                 |        |                         |                      |  |
|-------------------------------------|----------------------|-----------------|--------|-------------------------|----------------------|--|
| Culture<br>vessel                   | Surface<br>area/well | Fix<br>Solution | 1X PBS | AP Staining<br>Solution | Reactions<br>Per kit |  |
| 24-well<br>plate                    | 2 cm <sup>2</sup>    | 0.5 ml          | 0.5 ml | 0.6 ml                  | 50                   |  |
| 12-well<br>plate                    | 3.8 cm <sup>2</sup>  | 1 ml            | 1 ml   | 1 ml                    | 24                   |  |
| 6-well<br>plate                     | 9.6 cm <sup>2</sup>  | 2 ml            | 2 ml   | 1.5 ml                  | 12                   |  |

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